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Matthews, Barry ORCID logoORCID: <https://orcid.org/0000-0001-7605-0694> (2018) Pulse oximetry: SpO2 and SaO2. Journal of Paramedic Practice, 10 (3). pp. 126-128. doi:10.12968/jpar.2018.10.3.126

Official URL: <https://www.magonlinelibrary.com/doi/full/10.12968/jpar.2018.10.3.126>

DOI: 10.12968/jpar.2018.10.3.126

EPrint URI: <https://eprints.glos.ac.uk/id/eprint/8449>

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Pulse oximetry: SpO₂ and SaO₂

Barry Matthews

This article looks at the measurement of oxygen in the blood. It discusses how this essential observation is carried out in the pre-hospital environment, and its uses and limitations. The article will also take a brief look at the history and technology of the pulse oximeter.

History

Arterial oxygen saturation or the discovered oxygen level in arterial blood (SaO₂) is invasive, and it is difficult to monitor trends in a practice setting. In 1972, the idea of a non-invasive pulse oximeter to measure peripheral oxygen level (SpO₂) was conceived, with a prototype completed by 1974 (Aoygi, 2003). Prior to this, a number of monitoring methods for hypoxaemia have been described by Severinghaus (2011). Up until the 1930s, dentists were using pure N₂O (unlike the 50:50 N₂O:O₂ mix in Entonox®) to anaesthetise patients for teeth extraction. The cyanosis, or shade of blue of the patient, was used to monitor how long they could stay in this state. While the drugs improved in the 1930s, such as the use of thiopental, the monitoring did not. Respirations were impaired by the new exploration of anaesthetics, and the only change to monitoring was observing chest movement. Deaths inevitably followed.

In 1939, the oxyhemoglobinograph was developed by McClure and Hartman after the publication of their paper on anoxia during surgery (McClure et al, 1939), although this product was never made commercially available. The next iteration was a heavy device with an ear probe with the intention of monitoring aircraft pilots after World War II—the Millikan Oximeter. However, this was still too heavy and bulky to be a useful commercial product.

In 1947, there was an attempt at the measuring SpO₂ using the Millikan ear oximeter to consider the usefulness of cyanosis as a clinical indicator of hypoxia (Comroe and Bothelho, 1947). In 50% of patients, cyanosis did not occur until SaO₂ was below 80%; in 25% of patients, it occurred below 75% SaO₂. With this knowledge in hand, the recommendation of cyanosis as lone clinical indicator during anaesthesia would result in a high mortality rate. This brought on the pursuit of a commercially viable pulse oximeter.

The pulse oximeter

The principle behind the current approach towards pulse oximetry is that oxygenated blood absorbs light at a certain wavelength (a shade of red), and deoxygenated blood, another (infrared) (Nitzan, 2014). By shining these two light wavelengths through a peripheral area on a body and comparing the ratio of absorbencies against a calibration curve, the SpO₂ can be obtained. This is in turn used as an approximation for the patient's SaO₂.

A probe is used comprised of two parts sitting either side of the periphery: a light emitter, and a light receiver. The difference in colour between oxygenated and deoxygenated blood was commented on as early as 1864 (Stokes, 1864) as shown on the newly invented absorbance spectrometer by Kirchoff and Buncen (1860).

This estimation of SaO₂ by gaining the SpO₂ value from pulse oximetry is highly accurate, with a mean difference of less than 2% variation from SaO₂ samples over 90%. Below 90% SaO₂, this accuracy decreases (Jubran, 2015).

Despite these high-accuracy single-point estimates, changes in SaO₂ are not reliably predicted. If rapid changes towards hypoxaemia are expected, the probe attached to the earlobe may show these changes earlier than if used on a finger (Lindholm et al, 2007). This slow change is observable with a steady decline in SpO₂ on apnoea, compared with a sudden decline of EtCO₂.

Owing to the pulse and changes in blood flow, absorption varies throughout the pulse cycle. The changes in amplitude of the absorption not only allow for the pulse to be measured, but also for the distinction between venous and arterial blood, giving us the SpO₂ of arterial blood (Chan et al, 2013).

Application

As with all patient assessment, excellent hand and equipment hygiene (Bradley and Fraise, 2009) and an informed patient consent are of the utmost importance, with an explanation of the procedure and reasoning behind it.

- Select a site that is well perfused with a proximal palpable pulse; warm; with a brisk capillary refill; immobile; comfortable; and easily accessible (Welch et al, 2017)
- This is commonly either the fingers or earlobes, but other sites such as the tongue, cheeks, toes, or nose may be used in a patient with low peripheral perfusion (Barnett et al, 2012)
- Select an appropriately sized pulse oximeter probe so that it comfortably fits the patient (*Figure 1*). If it is too loose, the probe is likely to fall off; too tight, and venous pulsations may occur. This is an especially important consideration when using the adhesive probes—this type of probe may be more stable if not too tight

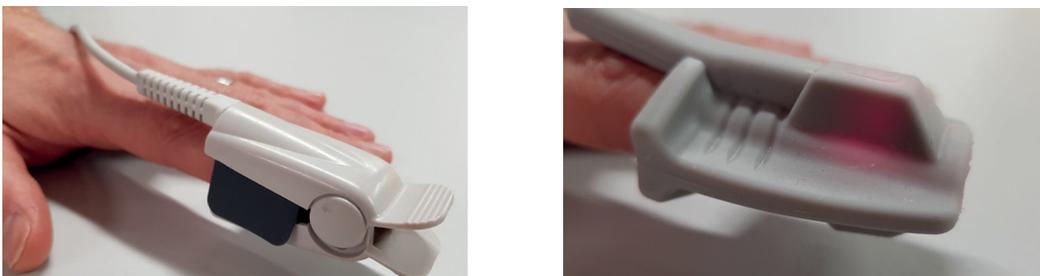


Figure 1. An appropriately sized pulse oximeter probe must be selected for comfort and access

- In paediatric patients weighing less than 3 kg, the ball of the foot can be considered; above this weight, the nail bed of the big toe is recommended (*Figure 2*). The same methods for pulse oximetry in adults can be adopted for older paediatric patients (Trigg and Mohammed, 2014). When using the toe, orientate the cable medially, and replace the sock to stabilise and ensure a strong signal

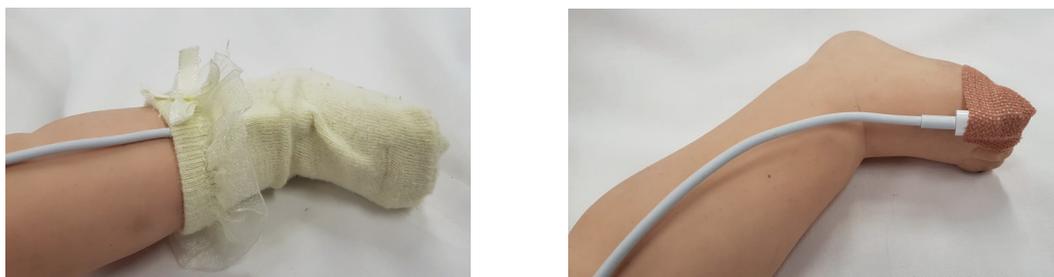


Figure 2. The nail bed of the big toe is recommended for paediatric patients weighing more than 3kg

- Remember that the probe measures the absorbency of pulsatile blood, so although a loose probe may have an artificially low SpO₂, venous pulsations from a tight probe or tape may also cause an artificially low SpO₂ by incorrectly measuring the venous blood (Chan et al, 2013)
- Consider environmental factors to reduce interference and improve signal strength. Electromagnetic radiation such as that produced by mobile phones can cause artefacts (Ortega et al, 2011)
- Patient or probe movements including muscular spasms, shivering, seizures or an inconsolable infant can also cause interference (Clarke et al, 2014).

Limitations

Despite the usefulness of this tool, there are limitations as a result of physiology, pathology or physical

changes. Changes to the colour of the haemoglobin, such as in the case of methemoglobin, which is blue/brown, can affect absorbency ratios. This form normally represents 2% of haemoglobin. In higher ratios, this has been shown to artificially maintain an SpO₂ of 85% in dogs, regardless of the correct SaO₂ (Barker et al, 1989). This can be genetically inherited, or caused by oxidising drugs such as nitrates.

In carbon monoxide poisoning, the irreversible carboxyhaemoglobin complex is formed, which is the same colour as oxyhaemoglobin. This will then not accurately reflect the SpO₂ as the absorbency ratio may be artificially altered (Fouzas et al, 2011).

Absorbance of the red light does not account for how freely the oxygen molecules can leave the haemoglobin link they have formed, and move to where they are required for aerobic respiration. Oxygen is more readily released when there is a drop in pH and an increase in carbon dioxide in the surrounding tissue, such as in muscles being used for physical activity having a higher cellular respiration rate.

If there were any concerns about a patient with a shifted dissociation curve, use of the probe on the ear lobe goes some way to counteract this over a finger placement, giving a truer reflection of central PO₂ (amount of oxygen dissolved in the blood away from haemoglobin that is available for cellular respiration) (Lindholm et al, 2007).

In patients with sickle cell crisis, evidence of hypoxaemia can be useful in determining the severity of the patient condition. SaO₂ can be underestimated with the SpO₂ reading gained by a pulse oximeter, showing accuracy only with an SpO₂ over 94% (Holbrook and Quinn, 2008; Zheng et al, 2016).

Nail polish or poor hygiene can affect how much light gets through. The light needs to be absorbed through the finger for the absorbance ratio to be produced. Nail varnish might seem like an obvious hindrance, but this has been challenged and there are arguments to both support and disregard the effect of the patient wearing nail varnish being clinically significant (Rodden et al, 2007). The finger (*Figure 3*) or whichever site chosen, may need to be cleaned prior to attempting the reading using the pulse oximeter.



Figure 3. A taped finger probe is another example of a pulse oximeter

When the SpO₂ reading reduces to below 80%, the comparability to SaO₂ starts to suffer. Feiner et al (2007) discovered up to a 3.6% mean bias for some brands of SpO₂ pulse oximeters, with a maximum discrepancy of 10% away from the actual SaO₂. The paper describes how pigmented skin can cause an over-estimate of SaO₂ in individuals with an SaO₂ of below 80%. Feiner et al (2007) also describes discrepancies based on gender.

Treat your patient

With all these limitations, the use of an SpO₂ pulse oximeter for the estimation of SaO₂ for a diagnosis of hypoxaemia should be used with caution. It can be seen that excessive hypoxia, or over-oxygenation in patients, can cause significant harm, both in the short and long term (Martin and Grocott, 2013). Reading the physical characteristics of the patient are more important than what can be read on the monitor— returning to the old dictum of ‘treat the patient, not the machine.’ JPP

References

- Aoyagi T. Pulse oximetry: Its invention, theory, and future. *J Anesth*. 2003;17(4):259-266.
- Barker SJ, Tremper KK, Hyatt J. Effects of methemoglobinemia on pulse oximetry and mixed venous oximetry. *Anesthesiology*. 1989;70(1):112–117.
- Barnett E, Duck A, Barraclough R. Effect of recording site on pulse oximetry readings. *Nurs Times*. 2012;108(1–2):22–23.
- Bradley C, Fraise AP. *Ayliffe's control of healthcare-associated infection fifth edition*. 5th edn. London: CRC Press; 2009.
- Clarke GWJ, Chan ADC and Adler A. Effects of motion artefact on the blood oxygen saturation estimate in pulse oximetry. *Medical measurements and applications (MeMeA)*. Ottawa, Canada. Piscataway: The Institute of Electrical and Electronics Engineers, Inc; 2014.
- Comroe JH, Botelho S. The unreliability of cyanosis in the recognition of arterial anoxemia. *Am J Med Sci*. 1947;124(1):1.
- Chan ED, Chan MM, Chan MM. Pulse oximetry: Understanding its basic principles facilitates appreciation of its limitations. *Respir Med*. 2013;107(6):789–799. <https://doi.org/10.1016/j.rmed.2013.02.00>
- Feiner JR, Severinghaus JW, Bickler PE. Dark skin decreases the accuracy of pulse oximeters at low oxygen saturation: The effects of oximeter probe type and gender. *Anesth Analg*. 2007;105(6 Suppl): S18–S23.
- Fouzas S, Priftis KN, Anthracopoulos MB. Pulse oximetry in pediatric practice. *Pediatrics*. 2011;128(4):740–752.
- Holbrook SP, Quinn A. An unusual explanation for low oxygen saturation. *Br J Anaesth* 2008;101(3):350–353.
- Jubran A. Pulse oximetry. *Critical Care*. 2015; 19:272.
- Kirchhoff G, Bunsen R. Professors bunsen and kirchhoff's method of chemical analysis by spectrum observations. *Annalen Der Physik Und Der Chemie*. 1860;110(1860):161–189.
- Lindholm P, Blogg SL, Gennser M. Pulse oximetry to detect hypoxemia during apnea: Comparison of finger and ear probes. *Aviation, Space and Environmental Med*. 2007;78(8):770–773.
- Martin DS, Grocott MPW. Oxygen therapy in critical illness: Precise control of arterial oxygenation and permissive hypoxemia. *Crit Care Med*. 2013;41(2):423.
- McClure RD, Hartman FW, Schnedorf JG, Schelling V. Anoxia: A source of possible complications in surgical anesthesia. *Ann Surg*. 1939;110(5):835–850.
- Nitzan M, Romem A, Koppel R. *Pulse oximetry: Fundamentals and technology update*. Auckland: Medical Devices. 2014;7(1):231–239.
- Ortega R, Hansen CJ, Elterman K, Woo A. Pulse oximetry. *New Engl J Med*. 2011;364(16): e33.
- Rodden AM, Spicer L, Diaz VA, Steyer TE. Does fingernail polish affect pulse oximeter readings? *Intensive Crit Care Nurs*. 2007;23(1):51–55.
- Severinghaus J. Monitoring oxygenation. *J Clin Monit Comput*. 2011;25(3):155–161.
- Stokes MA. The reduction and oxidation of the colouring matter of the blood. *Proc Roy Soc London*. 1864;13(1863–1864):355–364.
- Trigg E, Mohammed T. *Practices in children's nursing: Guidelines for hospital and community*. 3rd edn. London: Churchill Livingstone; 2010.
- Welch J, Osborne S, Adam S. *Critical care nursing: Science and practice*. 3rd edn. Oxford: Oxford University Press; 2017.
- Zheng S, Chan F, Ruiz G, Rees D, Gupta A. Pulse oximetry is an unreliable measure of haemoglobin oxygen saturation as calculated by earlobe blood gas co-oximetry in ambulatory paediatric sickle cell disease patients. *Arch Dis Childhood*. 2016;101(Suppl 1): A290.